

SOME ASPECTS ON THE HYDRATION OF SURFACTANT MICELLES

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Abstract - A review of the hydration properties of surfactant micelles is presented. Water self-diffusion studies are found to give global hydration numbers corresponding approximately to the hydration of the counterions and the polar head-groups. The hydration numbers are inconsistent with water penetration into micelles and other arguments against such an effect are discussed as well. Furthermore, the hydration of counterions and the effect of solubilization on hydration are discussed. Sodium cholate differs considerably in its self-association behaviour from micelle-forming amphiphiles and this is found to apply also for the hydration characteristics.

INTRODUCTION

The aggregation of amphiphilic compounds into micelles and other structures, being associated with eliminating the unfavourable contact between the non-polar groups and water, is an event specific to aqueous solutions. An important question refers to exactly how much of the amphiphile - water contact that is retained in the micelles. That the major part of the hydrocarbon chain - water contact is eliminated on micelle formation is inherent in our view of the driving forces of micelle formation but has also been directly demonstrated by a large number of experimental studies. However, several authors have presented strikingly opposing views of micelle hydration. These are mainly based on spectroscopic probe studies which have been taken to indicate that there is an extensive penetration of water far into micelles. The present article, in reviewing previous results and presenting some novel experimental information, examines in a broad way the matter of the hydration of amphiphilic aggregates. Questions to be discussed include global hydration numbers, location of water molecules, counterion hydration, hydration dynamics and effect of solubilization on hydration. In particular we examine the evidence taken to show that there is a deep water penetration into micelles.

In considering the exposure of the hydrocarbon chains to water, the dynamic state of the micelle must be taken into account. The life-time of a monomer in a micelle may be of the order of a microsecond and, in view of the well-established dynamic state of a micelle, this implies that less extensive motions occur on a shorter time-scale. The dynamic protrusion of methylene groups from the hydrocarbon core of a micelle has recently been examined by Aniansson (1) who came to the conclusion that there is a considerable protrusion. Thus every third monomer would protrude more than one methylene group and the average protrusion would be one methylene group. A considerable roughness of the micellar surface has been inferred also by others and an attempt to visualize the dynamic roughness of the micelle surface is given in Fig. 1.

GLOBAL HYDRATION NUMBERS OF MICELLES

There is a large number of both thermodynamic, transport and spectroscopic methods which can be used for investigations of the hydration of amphiphilic aggregates. The micelle-water interaction is highly dynamic and stoichiometrically not well defined and, therefore, the concept of a single hydration number is a simplification; the meaning of a hydration number will be dependent on the particular experimental approach considered. An initial suitable definition for our present purpose is to take the micelle hydration number as the number of water molecules moving with the micelle as a kinetic entity. Such a global hydration number can be deduced from transport properties, e.g. viscosity and diffusion. Mukerjee (2) has given a useful description of the evaluation of hydration numbers from viscosity data. The procedure involves

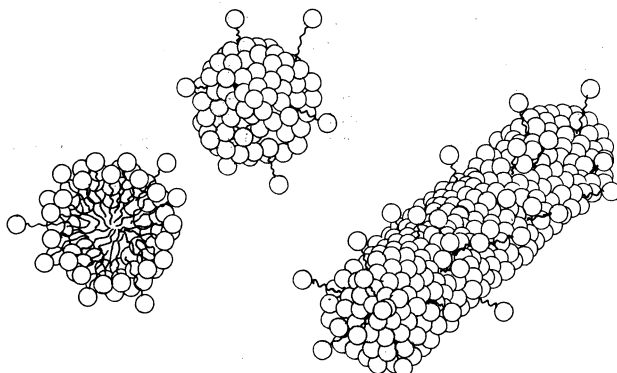


Fig. 1. A picture of spherical and cylindrical micelles showing the protrusion of some amphiphile molecules. The interior shows the disordered state of the hydrocarbon chains. (By the courtesy of J. Ulmius.)

the determination of the intrinsic viscosity and comparing it with the partial specific volume of the amphiphile. The precision of the method is reduced since certain corrections, mainly for electroviscous effects, may have to be applied.

Ourselves, we have been interested in determining micelle hydration numbers from the translational self-diffusion coefficient of water. Ideally, some of the water molecules can be considered as associated with the micelles and having the translational mobility of the micelles while the rest of the water molecules are free having the translational mobility of pure water. In reality, certain corrections may have to be applied, notably for the hydration of the non-associated amphiphile and of the free counterions and furthermore, the mobility of free molecules may be retarded *inter alia* due to obstruction effects. The observed diffusion coefficient can be written as an average over all the different environments i that the water molecules sample, i.e.

$$D = \sum P_i D_i \quad (1)$$

where P_i is the fraction of the water molecules in site i characterized by the diffusion coefficient D_i . We used as an experimental approach the open-ended capillary tube method employing tritium labelling. Hereby it is usually possible to determine diffusion coefficients with a precision of 1-2 %. However, the applicability of the technique depends on the consistency etc. of the sample; for highly viscous cases, studies are thus precluded and for anisotropic phases the interpretation is difficult.

It is a general observation in our studies of the water diffusion as a function of the surfactant concentration that the diffusion coefficient decreases much more rapidly below than above the c.m.c. The mobility of a water molecule bound to a micelle is much lower than that of a water molecule hydrating the surfactant monomer but the reduction in surfactant-water contact is evidently enough to compensate for this effect. It is possible to quantify the effect using eq. (1) which can schematically be written

$$D_w = p_w^{nm} D_w^{nm} + \frac{n_{mic} m_{mic}}{55.5} D_{mic} \quad (2)$$

where D_{mic} is the diffusion coefficient of the micelles, m_{mic} is the molality of surfactant in micellar form and n_{mic} the micelle hydration number (per amphiphile ion). p_w^{nm} is the fraction of water molecules not associated with the micelles and D_w^{nm} the corresponding effective diffusion coefficient. In reality the first w term of eq. (2) consists of three parts corresponding

in addition to the free water molecules to the water molecules hydrating the free amphiphile ions and the free counterions. The magnitude of this term can be established knowing the concentrations of free amphiphile ions and counterions, their self-diffusion coefficients and the effect of the non-associated surfactant on the water self-diffusion coefficient (from pre-c.m.c. studies). In the second term, D_{mic} can be determined separately (for example from the self-diffusion coefficient of a solubilizate (3)) but in most cases $D_{mic} \ll D_w^{nm}$ so that the second term may be neglected. The main effect of micelle hydration is then a reduction of p_w^{nm} , a reduction which is due to micelle hydration and gives information on n_w^{mic} . A further simplification is often possible for surfactants with low c.m.c.s in that the correction for a change in D_w due to an altered concentration of non-associated surfactant is negligibly small.

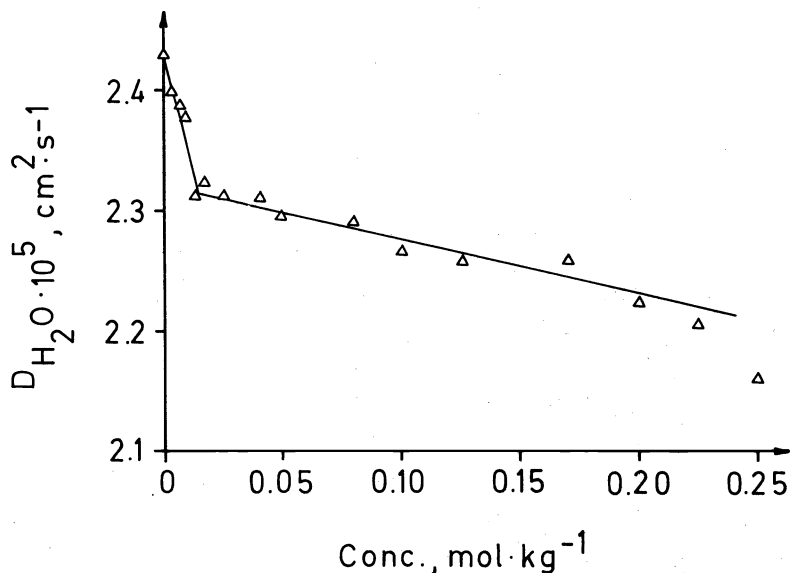


Fig. 2. Concentration dependence of the water self-diffusion coefficient for solutions of sodium octylbenzenesulfonate at 33 °C. (By the courtesy of M.-C. Puyal.)

An example of a determination of micelle hydration by the self-diffusion method is given in Fig. 2 for the case of sodium p-octylbenzenesulfonate at 33 °C. Evidently, the concentration dependence of D_w can be described by two linear segments intersecting at the c.m.c. The hydration number obtained is ca. 9 and as can be seen in Table 1, rather similar results are obtained for some other surfactants. Mukerjee (2, 4) some time ago concluded from this type of data for a few cases that the micellar hydration numbers can be accounted for in terms of hydration of the polar head-groups and the counterions. As can be seen, the more recent self-diffusion results support this view and as will be demonstrated below it appears that many simple counterions retain their primary hydration sheaths on binding to micelles.

WATER PENETRATION INTO MICELLES

According to the classical picture of surfactant micelles, as it was first clearly presented by Hartley in the 1930's, there is in the micelle a mini-mization of the contact between the alkyl chains and the water. It is now well established that the main driving force for micellization is the hydrophobic interaction (Refs. 5 and 6 are recent reviews of the subject). This picture of micelles has been questioned at several occasions mainly in connection with spectroscopic studies. Thus it has been suggested that there is an important penetration of water deep into micelles. Earlier results of this type (7 - 9) were strongly refuted by Mukerjee (4, 10) and Stigter (11) but as the matter has again been advanced by Menger (12 - 14) in some recent papers it deserves discussion.

TABLE 1. Micelle hydration numbers expressed as the number of water molecules per amphiphile ion.

<u>Amphiphile</u> ^a	<u>Hydration number</u>
C ₁₂ OSO ₃ Na	9 ^b
C ₁₂ OSO ₃ Na	8 ^c
C ₁₂ OSO ₃ Na	12 ^d
C ₁₂ N(CH ₃) ₃ Cl	5 ^b
C ₁₄ N(CH ₃) ₃ Cl	5 ^b
C ₇ CO ₂ Na	8.5-8.9 ^e
C ₇ CO ₂ Na	8.7 ^f
C ₈ (C ₆ H ₄)SO ₃ Na	9 ^g
C ₁₂ NO(CH ₃) ₂	10 ^d
C ₈ NO(CH ₃) ₂	10 ^d

a. C_n is a n-alkyl chain with n carbon atoms.

b. Ref. 2.

c. F. Tokiwa and K. Ohki, *J. Phys. Chem.* 71, 1343-1348 (1967).

d. W. L. Courchene, *J. Phys. Chem.* 68, 1870-1874 (1964).

e. P. Ekwall and P. Holmberg, *Acta Chem. Scand.* 19, 455-468 (1965).

f. Ref. 25.

g. M.-C. Puyal, D.E.A., USTL, Montpellier, 1978.

In a typical spectroscopic experiment concerning the amphiphile-water contact, one monitors a parameter of a probe, intrinsic or added, and correlates the value with that found in a pure solvent of a given polarity. Such an approach is connected with two problems, the seriousness of which is often overlooked. Firstly, to be able to deduce information about the molecular environment in the micelle one must have independent information on the distribution of the probe in the micelle. Secondly, the relation between the spectroscopic parameter in question and the nature of the molecular environment of the probe should either have a sound theoretical basis or it should be experimentally tested in detail.

Several studies giving a water penetration into micelles may be criticized on the basis of one or both of these points. A typical example is provided by the careful ¹⁹F NMR chemical shift studies by Muller and co-workers (7, 8). These authors inter alia investigated ω-CF₃ substituted amphiphiles. Hereby it was found that the ¹⁹F chemical shift is approximately midway of that of water and hydrocarbon solutions. This could be taken to imply that the -CF₃ group has an environment consisting of about equal amounts of water and hydrocarbon (7, 8) but as shown recently by Mukerjee (4, 10) and by Muller (15) such an interpretation can be criticized on the basis of the two points mentioned above. Mukerjee (4, 10, 16) gave strong arguments for an important sampling of the -CF₃ groups of the micelle surface. Thus there is a marked non-ideality of the interactions between fluorocarbons and hydrocarbons which is manifested for micelles in terms of anomalously high c.m.c.s for partially fluorinated surfactants and a partial demixing of the micelles for solutions containing both fluorocarbon and hydrocarbon surfactants. Muller (15) has recently shown in a detailed study of the factors influencing ¹⁹F chemical shifts that the underlying mechanism is too complex to permit conclusions about environment polarity.

Menger et al. (12) studied water penetration using the amphiphile

CH₃(CH₂)₇ $\overset{\text{O}}{\parallel}\text{C}$ (CH₂)₇N⁺(CH₃)₃Br⁻ correlating the ¹³C NMR chemical shift of the

carbonyl carbon with the shifts obtained in a range of solvents. Again one does not have a sufficiently good theoretical basis of the molecular cause of the shifts nor does one have independent information on the distribution of the carbonyl group in the micelle. The use of a polar probe like - $\overset{\text{O}}{\parallel}\text{C}$ - can certainly not be expected to report reliably on the environment $\overset{\text{O}}{\parallel}\text{C}$

of the alkyl chains in a micelle. Rodgers and da Silva E Wheeler (17) investigated the quenching of pyrene fluorescence by inorganic ions and interpreted the effective quenching in terms of an extensive water penetration deep into micelles. An assumption inherent in this work was though that pyrene is solubilized remote from the micellar surface. However, it seems that there is very good evidence for the solubilization of aromatic compounds close the head-group region (4, 18 - 20) and there is, therefore, no need to invoke any water penetration to explain these data. As regards a Raman spectroscopic study (21) taken to support a less extensive water penetration it can be said that the mode of interpretation has no theoretical basis but is based on analogy. This refers also to a study of the vibronic fine structure in pyrene fluorescence (22), but here pyrene solubilization was, furthermore, taken to be in the interior of the micelles rather than in the surface region, as in more recent studies (23, 24). Svens and Rosenholm (9) from x-ray scattering data inferred partial water penetration into micelles but it is clear that the model used in the analysis is oversimplified. Menger (12 - 14) cites this work in favour of a much deeper water penetration than implied in the original paper (9).

It seems thus clear that the results taken as support of water penetration have alternative reasonable interpretations and that there, therefore, is no reason to abandon the classical picture of a micelle as an aggregate with an apolar interior and a polar surface in contact with the surrounding aqueous medium (4, 11). A most significant argument against a deep water penetration seems to be given by the global hydration numbers of micelles (2, 11, 25). The marked slope reduction at the c.m.c. in Fig. 2 is a very direct demonstration that a strongly dominating part of the amphiphile-water contact is eliminated on micellization. Several other pertinent observations also exist. Brun et al. (26) obtained large positive isentropic apparent compressibilities of several surfactants in micelles which indicates that the interior of micelles resembles liquid hydrocarbons. Furthermore, the partial molar volumes and compressibilities of solubilized alkanes are close to the values of liquid alkanes but considerably higher than those in water (27). The H/D isotope effect on ^{19}F NMR relaxation for partially fluorinated and fluorocarbon surfactants disappears on micelle formation (28). Podo et al. (29) observed no H/D isotope effect on ^1H NMR relaxation of the alkyl protons, but a noticeable one for the ethoxy ones, for nonionic surfactant micelles.

The low solubility of water in alkanes is of course in support of the absence of water penetration and more direct evidence on this point can be obtained making use of the established similarity between different surfactant-water phases. Thus there is a low rate of water diffusion between reversed micelles (30) as well as over lipid bilayers (31). Furthermore, the stability ranges of different phases (32) are inconsistent with any marked water penetration. Neutron diffraction studies using the $\text{D}_2\text{O}/\text{H}_2\text{O}$ exchange method show that water penetrates to the glycerol-fatty acid ester group for phospholipid bilayers (33 - 35).

Menger (13) has recently from the building of space-filling molecular models argued that it is impossible to pack the surfactant molecules into micelles without leaving sizeable voids filled with water. However, this does not agree with work by Gustavsson (cited in Ref. 5) which demonstrates the feasibility of packing surfactant monomers into micelles without any water contact except at the micellar surface.

SOME FURTHER POINTS ON THE HYDRATION OF AMPHIPHILIC AGGREGATES

Although the general features of micelle hydration, with a limited number of water molecules located mainly at the micellar surface, thus seem well supported, it is of interest also to consider some additional aspects. For example it is important to know to what extent bound counterions retain their hydration sheath. For alkali counterions, there is significant evidence to suggest that the primary hydration sheath is retained. This has been demonstrated from volumetric studies (36, 37) and NMR studies which monitor directly the counterion give good support for this. Thus $^{23}\text{Na}^+$ relaxation in surfactant systems remains slow down to quite low water contents (38) and $^{23}\text{Na}^+$ and $^{133}\text{Cs}^+$ chemical shift changes on micelle formation are quite small in comparison with what would be expected for direct counterion-head-group approaches (39). An illustrative observation, reproduced in Figure 3, is that the H/D water isotope effect in $^{133}\text{Cs}^+$ chemical shift does not

change even at very high concentrations (39).

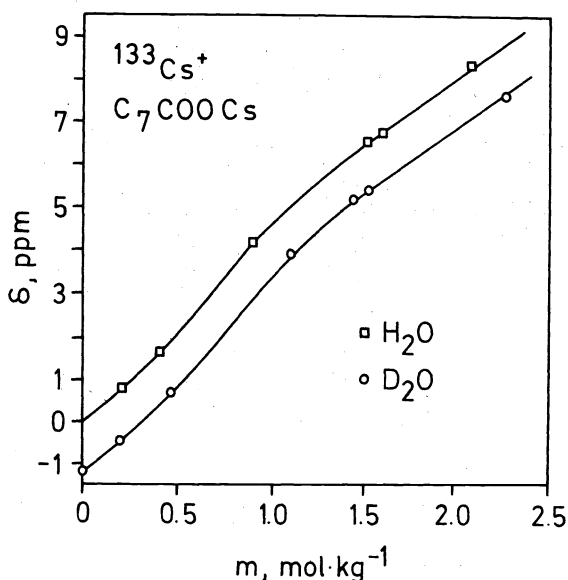


Fig. 3. ^{133}Cs chemical shifts (ppm) of H_2O and D_2O solutions of cesium octanoate as a function of amphiphile molality. (After Ref. 39.)

Concerning the halide ions, we have yet incomplete information. The observation of charge-transfer interactions for I^- (40) and of rather large changes in Cl^- and Br^- quadrupole relaxation rates (41) on binding to micelles may be taken to suggest that there are significant direct counterion-head-group approaches. The marked counterion specificity noted in some cases for c.m.c., micellar size etc. points in the same direction.

Deuteron NMR quadrupole splittings of D_2O have been found useful to monitor the water orientation at amphiphilic surfaces in anisotropic surfactant-water phases, like lamellar and hexagonal mesophases. Since for a number of systems only ca. 5 water molecules per amphiphile are appreciably oriented and the water orientation is markedly alkali counterion dependent (42), these studies support the view of a limited hydration number and of hydrated counterions.

How solubilization influences micelle hydration is rather difficult to study since changes in physico-chemical parameters are small. For approximately spherical micelles of hexadecyltrimethylammonium bromide, self-diffusion studies indicate some release of hydration water on solubilization of octanol and hexanol but not on solubilization of cyclohexane (43); for higher concentrations there were interpretation difficulties. For anisotropic phases, the ^2H quadrupole splitting method can be useful. A study of hexagonal phases of water and sodium octanoate or sodium octylsulfate gave pronounced changes in the ^2H quadrupole splitting of D_2O (44). Roughly, a more polar solubilizee decreases water binding while a non-polar one gives a small effect or an increased water binding. This is according to expectation since a compound solubilized close to the micelle surface may displace some water whereas a compound solubilized in the micellar interior may increase the surface per polar group and thus increase hydration to some extent.

As mentioned above, it is well established that the micelle is in a highly dynamic state. A recent ^{13}C NMR study showed that there is no marked change in the alkyl chain mobility on micellization (45) and likewise the water molecules of micelle hydration are very mobile. Although quantitative information is lacking it may be suggested that the motion has a time-scale of the order of 10^{-11} - 10^{-12} sec.; the local motion at the amphiphile aggregate surface is not far from being isotropic as can be inferred from the

low value of the order parameter (ca. 0.01) of the bound water molecules (42). As regards water exchange kinetics, we know from the NMR studies that there is a rapid water exchange but quantitative information has not been available. However, Tiddy et al. (46) recently observed an ultrasonic relaxation in the frequency range around 10 MHz for concentrated surfactant solutions and mesophases which they attributed to water exchange; further studies along these lines will be followed with great interest.

This discussion has concerned the hydration of aggregates of typical micelle-forming amphiphiles but it may be of interest for comparison to consider results for an amphiphile of another type, sodium cholate. It has been demonstrated that sodium cholate is not characterized by the strongly cooperative association into large aggregates which occurs for surfactants (47 - 50). Our self-diffusion studies further illustrate the marked difference in several respects between sodium cholate and micelle forming surfactants (51, 52). Thus the fraction of cholate aggregated changes only slowly with concentration and the aggregates remain fairly small over a wide concentration range. The degree of counterion binding is highly variable being quite low for the first-formed aggregates but increasing progressively to values typical of surfactants at high concentrations. For micelle-forming amphiphiles, the degree of counterion binding generally has a closely constant value over a wide concentration range. As can be inferred from Fig. 4, hydration of

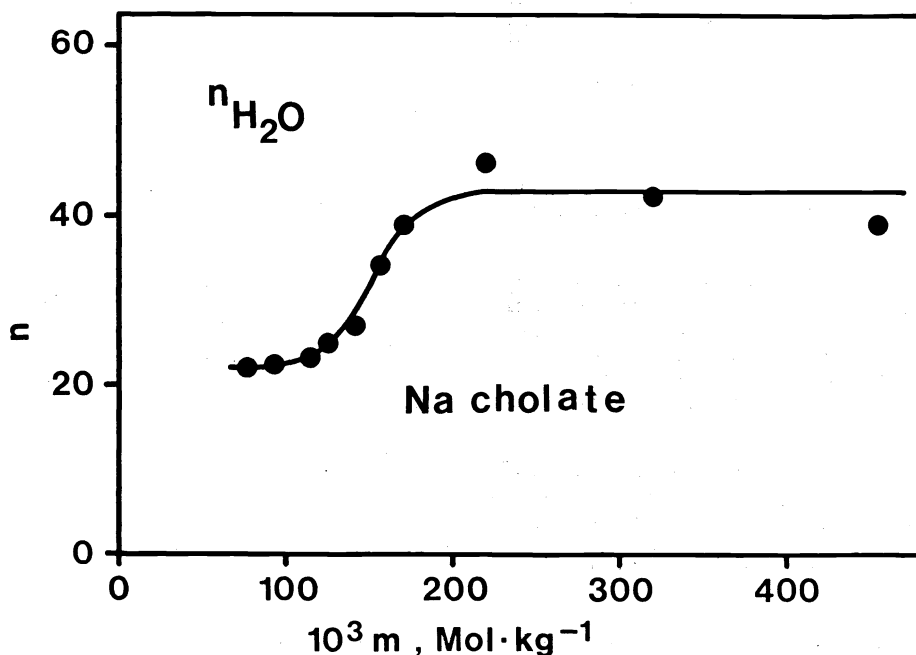


Fig. 4. Hydration number of cholate aggregates as a function of the sodium cholate molality according to self-diffusion studies. (After Ref. 52.)

cholate aggregates also shows some peculiarities. Firstly, the hydration number is high and, secondly, it increases with increasing concentration. (Certain difficulties in the evaluation necessitate some reservation as regards the exact magnitude of the effects.) For surfactant micelles, although not documented in detail, it seems that the hydration number stays essentially constant or shows some decrease at higher concentrations. Although far-reaching conclusions may not be drawn from these data they seem to provide good support for a model involving a primary association involving mainly hydrophobic interactions and a secondary association involving non-hydrophobic (hydrogen bond) interactions (cf. Refs. 50, 53 and 54).

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